CASE REPORT



A novel 14q13.1–21.1 deletion identified by CNV-Seq in a patient with brain-lungthyroid syndrome, tooth agenesis and immunodeficiency



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Abstract

Background: Chromosome 14q11-q22 deletion syndrome (OMIM 613457) is a rare genomic disorder. The phenotype heterogeneity depends on the deletion size, breakpoints and genes deleted. Critical genes like *FOXG1*, *NKX2*–1, *PAX9* were identified.

Case presentation: We performed whole exome sequencing (WES) and copy number variation sequencing (CNV-seq) for a patient with mild speech and motor developmental delay, short stature, recurrent pulmonary infections, tooth agenesis and triad of brain-lung-thyroid syndrome. By using CNV-seq, we identified a 3.1 Mb de novo interstitial deletion of the 14q13.2q21.1 region encompassing 17 OMIM genes including *NKX2–1, PAX9* and *NFKBIA*. Our patient's phenotype is consistent with other published 14q13 deletion patients.

Conclusion: Our results showed the combination of WES and CNV-seq is an effective diagnostic strategy for patients with genetic or genomic disorders. After reviewing published patients, we also proposed a new critical region for 14q13 deletion syndrome with is a more benign disorder compared to 14q11-q22 deletion syndrome.

Keywords: 14q13 deletion, CNV-seq, Brain-lung-thyroid syndrome, Immunodeficiency

Background

Chromosome 14q11-q22 deletion syndrome (OMIM 613457) is a genomic disorder characterized by microcephaly, dysmorphic facies, psychomotor delay and failure to thrive. The associated phenotype is heterogeneous, depending on the size and variable breakpoints [1–3]. Some major features can be explained by haploinsufficiency of critical genes like *FOXG1*, *NKX2*–1, *PAX9*, etc. [4, 5]. Patients with interstitial deletions involving 14q13.1q21.1 are rarely reported and all these patients had similar features. *NKX2*–1 and *PAX9* in 14q13 are

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considered to be the critical genes causing brain-lungthyroid syndrome (BLTS, OMIM 610978) and tooth agenesis (OMIM 604625) features, respectively [6, 7]. More recently, *NFKBIA* was purposed to be responsible for immunodeficiency these patients [8].

In our study, we report a new patient with 14q13.1q21.1 distal microdeletion syndrome. Copy number variation sequencing (CNV-seq) revealed a de novo 3.10 Mb sized deletion (chr14: 35,268,524–38,367,321). We detailed described this patient's phenotype and reviewed all reported patients with similar breakpoints encompassing *NFKBIA*, *NKX2*–1 and *PAX9*, and provided more information of relationships between clinical features and deleted genes.

Case presentation

The patient is a 15-year-old female born to nonconsanguineous parents by nature labor after full-term gestation. Family history is unremarkable. She was diagnosed



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as aspiration pneumonia at birth and stayed in hospital for 4 days. Developmental delay was noticed after birth. She could not sit until 12 months. She could speak after 24 months and walk after 27 months. Dysarthria and ataxia were also unnoticed since after. At the age of 2 years, she went to hospital for speech and motor developmental delay. The symptoms were relieved after rehabilitation training. She was also diagnosed as short stature when she was 14 and received growth hormone therapy for 9 months. Her height increased from 132 cm (- 4.7 SD) to 142 cm (- 3.2 SD), resulting in an annual growth velocity of 13 cm.

At the age of 15 years, she was referred to the Department of Respiration, Beijing Children's Hospital for continuous cough and expectoration for 3 years. Her height and weight were below 3rd percentile. She did not have obvious facial anomalies but hypohidrosis and tooth agenesis (missing secondary dentition including molars and premolars) were noticed (Fig. 1a-c). She also had punctate pigmentation was noticed on her back. Mental development was delayed compared to peer children. Immune system examination revealed decreased IgG (3.69 g/L), other immunoglobulins and lymphocyte subsets were normal. Paranasal sinusitis and anemia were diagnosed. She had increased thyroid-stimulating hormone (TSH, 11.983 mIU/L), deceased triiodothyronine (T3, 58.79 ng/dL) and normal tetraiodothyronine (T4, 4.81 μ g/dL), indicating a compensated hypothyroidism. She also had delayed development of the secondary sexual characters. Till this visit, she did not have menarche when she was 15. Her bone age was also delayed (BA = 10).

Thoracic computed tomography (CT) revealed interstitial and parenchymal lesions and bronchiectasis in both lungs, dominated with interstitial lesions, and mucus plugs was found in right upper lobe and left lower lobe (Fig. 1d). Ultrasonography of knee joint showed very limited amount of effusion in bilateral suprapatellar capsule. Routine blood test, coagulation function, and screening for infectious diseases, abdominal ultrasonography, ultrasonic cardiography, cranial CT and paranasal sinus CT were all normal.

DNA was isolated from peripheral blood samples obtained from the proband and her parents by using Gentra Puregene Blood Kit (QIAGEN, Hilden, Germany). 200 ng genomic DNA of each individual was sheared by Biorupter (Diagenode, Belgium) to acquire 150~200 bp fragments. The ends of DNA fragment were repaired and Illumina Adaptor was added (Fast Library Prep Kit, iGeneTech, Beijing, China). After sequencing library



were constructed, the whole exome was captured with AIExome Enrichment Kit V1 (iGeneTech, Beijing, China) and sequenced on Illumina NovaSeg 6000 (Illumina, San Diego, CA) with with 150 base paired-end reads. Raw reads were filtered to remove low quality reads by using FastQC. Clean reads were mapped to the reference genome GRCh37. Single nucleotide variants (SNVs) were annotated and filtered by TGex (http://app. genecards.cn) and classified following the American College of Medical Genetics and Genomics and the Association for Molecular Pathology interpretation standards and guidelines [9]. To identify large copy number variations (CNVs), part of the sequencing library was sequenced directly on Illumina NovaSeq 6000 and each sample yielded one Gigabase Raw data. An in-house pipeline was applied to map and call CNVs and the health parents were used as control samples [10]. Clean reads were mapped to the reference genome GRCh37. Database of Genomic Variants, DECIPHER database, ClinVar, OMIM and ClinGen were used for interpretation and classification of the clinical significance of candidate CNVs according to previously reported guidelines [11]. By whole exome sequencing (WES), 30,636 SNVs and small indels were called from 14,549.59 Mb Clean bases (Target mean depth was 122.23X) in the proband. After analysis and interpretation, none of these variants could explain the proband's phenotype. However, CNVseg revealed a de novo 3.1 Mb deletion on 14g13.1g21.1 (chr14: 35,268,524-38,367,321) (Fig. 2). The deletion encompasses 17 OMIM genes including BAZ1A, SRP54, PPP2R3C, KIAA0391, PSMA6, NFKBIA, INSM2, RAL-GAPA1, PTCSC3, MBIP, SFTA3, NKX2-1, NKX2-8, PAX9, SLC25A21, MIPOL1 and FOXA1. The deletion was not present in the parents. WES reads depth data was used to validate this deletion ($\log 2 = -1.00428$, mean depth = 50.02X, probes number = 379). Informed consent was obtained from the parents of the patient.

Discussion and conclusions

In this study, we described clinical and molecular findings in a female with mild speech and motor developmental delay, short stature, recurrent pulmonary infections, tooth agenesis and triad of BLTS. Using CNV-seq, we identified a 3.1 Mb de novo interstitial deletion of the 14q13.2q21.1 region encompassing 17 OMIM genes.

The prevalence of chromosome 14q11-q22 deletion syndrome is lower than 1 in 1,000,000 infancies. The phenotype of patients is largely determined by the deletion size and breakpoints. Several candidate genes were well identified like FOXG1, NKX2-1 and PAX9. Deletions including FOXG1 and a 1 Mb upstream region on 14q12 cause severe intellectual disability [5, 12–16]. NKX2-1 (14q13.3) deletion is responsible for choreoathetosis, hypothyroidism, short stature and neonatal respiratory distress and haploinsufficiency of PAX9 (14q13.3) causes oligodontia phenotype [5, 17–21]. The role of other genes is still under evaluation. NPAS3 (14q13.1) encodes a transcription factor localized to the nucleus and may regulate genes involved in neurogenesis. Npas3^{-/-} mice had abnormal neurodevelopment, neurosignaling and behavior [22], making it a candidate gene of holoprosencephaly (HPE) and hypoplasia of the corpus callosum (ACC) in 14q11-q22 deletion syndrome patients [5, 23]. RALGAPA1 (14q13.2) encodes a major subunit of the RAL-GTPase activating protein, and was





suggested to be important in brain development [24]. NFKBIA (14q13.2) mutation cause dominant inherited ectodermal dysplasia and immunodeficiency 2 (EDAID2, OMIM 612132) and may be the explanations of patients' immunological features [4, 8]. SEC23A (14q21.1) encodes a subunit of a protein complex and found in the ribosome-free transitional face of the endoplasmic reticulum and associated vesicle and it is considered as a candidate gene of joint hyperlaxity [8, 17]. Mice with knock-out had abnormal cartilage development and collagen level [25]. Biallelic SEC23A mutation causes craniolenticulosutural dysplasia (OMIM 607812). It is characterized by facial dysmorphism, late-closing fontanels, cataract, and skeletal defects including joint laxity [26].

The deletion region of our patient (chr14: 35,268,524-38,367,321) encompasses RALGAPA1 and NFKBIA but no NPAS3 or SEC23A. Consequently, joint laxity, HPE or ACC was not observed in our patient. Epilepsy was also absent in our patient even though RALGAPA1 was deleted. We reviewed published ten patients with phenotype description and similar deletion regions encompassing RALGAPA1, the deleted sizes ranged from 0.82 Mb to 6.98 Mb, and only one patient with 1.99 Mb deletion reported by Caliebe A et al. had seizures [4, 5, 8, 19, 23, 27-29]. Therefore, deletion of RALGAPA1 was not sufficient to cause seizures and the genotype-phenotype correlation of RALGAPA1 deletion remained unclear. Considering our patient had decreased IgG, paranasal sinusitis and recurrent infections, we also reviewed immunological features of seven patients with NFKBIA deletion (two patients were collected from DECIPHER database, Fig. 3). Patient reported by Villafuerte B et al. also had low IgG but also relatively low lymphocyte count and percentage of switched B cells [8]. Gentile M patient also had recurrent infections, with a mild reduction of CD3/CD8 lymphocytes and an elevation of CD4/

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CD8 ratio, yet her IgA, IgG, and IgM were normal [4]. Santen G patient 5 and Peall K patient 4 had recurrent lower respiratory infections [5, 19]. Patient 256,879 from DECIPHER database also had recurrent infections. We next reviewed seven patients with PAX9, NKX2-1 deletion and leaving NFKBIA intact in previously literature. The deleted sizes ranged from 0.36 Mb to 3.69 Mb, and only one male with 2.34 Mb had recurrent bronchitises [5, 17, 19, 20, 29]. It was notable that the pLI (probability of LoF intolerant) value of NFKBIA was 1, and the o/ e score is 0 (0-0.19), indicating deletion of this gene may have serious clinical consequences. In addition, three NFKBIA nonsense variants were reported in patients with EDAID2 [30]. To this respect, NFKBIA haploinsufficiency may be an appropriate explanation of the immunological features of 14q13 deletion patients.

Our patient, together with previously reported patients, identified a well-defined, more benign, 14q13 distal microdeletion syndrome. The major phenotype includes choreoathetosis, tooth agenesis, pulmonary dysfunction, immunological abnormal and hypothyroidism. The 1.4 Mb critical region contained at least three candidate genes: NFKBIA, NKX2-1 and PAX9 (Fig. 3). In this region, one more gene, PSMA6, with its pLI being 1 and o/e score being 0 (0-0.23), should be more attention. PSMA6 encodes the component of the 20S core proteasome complex involved in the proteolytic degradation of most intracellular proteins. Variants in PSMA6 were reported to be associated with inflammation diseases like myocardial infarction [31], arthritis [32, 33], etc. Given its essential function and intolerant of LoF variants in population, more subtle features may be uncovered in the future.

Previous researches have demonstrated that CNV occurred in 5–10% of the total human genome [34], and chromosomal microarray analysis (CMA), as a stable and accurate platform, is used for detecting. Currently,



CNV-seg is developed by analyzing data generated from WES or whole genome sequencing (WGS) [35-37]. Recent studies showed that the combination of WES and CNV-seq by low cover genome sequencing increased diagnostic yield in patients with rare diseases [38, 39]. In our study, after suspecting the diagnosis of BLTS of our patient, we performed trio-WES and low coverage WGS (0.3X) simultaneously. No putative pathogenic variants in NKX2-1 was identified, but we uncovered a 3.1 Mb deletion encompassing NKX2-1, as well as PAX9 and NFKBIA. Notably, WES data could be applied to CNV identification, so we used WES data to validate this deletion. By reviewing our deleted region and other reported patients, we proposed a 1.4 Mb critical region for 14q13 distal microdeletion syndrome with a well-defined, more benign phenotype compared to 14q11-q22 deletion syndrome. Our results further demonstrated the clinical utility of the diagnostic strategy combining WES and CNV-seq for genetic diseases patients.

Abbreviations

ACC: Hypoplasia of the corpus callosum; BLTS: Brain-lung-thyroid syndrome; CMA: Chromosomal microarray analysis; CNVs: Copy number variations; CNVseq: Copy number variation sequencing; CT: Computed tomography; HPE: Holoprosencephaly; pLI: Probability of LoF intolerant; SNVs: Single nucleotide variants; WES: Whole exome sequencing; WGS: Whole genome sequencing

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Authors' contributions

XH drafted the manuscript and analyzed WES and CNV-seq data. RG and JG performed molecular experiments. JL and ZZ performed the clinical examination of the patient and his family. WL performed data interpretation and reviewed the manuscript. BX and CH managed the patient and critically revised the paper. All authors read and approved the final manuscript.

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Availability of data and materials

The datasets used and/or analyzed during the current study are available from the corresponding author on reasonable request.

Ethics approval and consent to participate

The study was approved by the Ethics Committee of Beijing Children's Hospital, Capital Medical University.

Consent for publication

Informed consent was obtained from the parents of the patient.

Competing interests

The authors declare that they have no competing interests.

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References

- Grammatico P, de Sanctis S, di Rosa C, Cupilari F, del Porto G. First case of deletion 14q11.2q13: clinical phenotype. Ann Genet. 1994;37(1):30–2.
- Kamnasaran D, O'Brien PC, Schuffenhauer S, Quarrell O, Lupski JR, Grammatico P, et al. Defining the breakpoints of proximal chromosome 14q rearrangements in nine patients using flow-sorted chromosomes. Am J Med Genet. 2001;102(2):173–82.
- Torgyekes E, Shanske AL, Anyane-Yeboa K, Nahum O, Pirzadeh S, Blumfield E, et al. The proximal chromosome 14q microdeletion syndrome: delineation of the phenotype using high resolution SNP oligonucleotide microarray analysis (SOMA) and review of the literature. Am J Med Genet A. 2011;155A(8):1884–96.
- Gentile M, De Mattia D, Pansini A, Schettini F, Buonadonna AL, Capozza M, et al. 14q13 distal microdeletion encompassing NKX2-1 and PAX9: patient report and refinement of the associated phenotype. Am J Med Genet A. 2016;170(7):1884–8.
- Santen GW, Sun Y, Gijsbers AC, Carre A, Holvoet M, Haeringen A, et al. Further delineation of the phenotype of chromosome 14q13 deletions: (positional) involvement of FOXG1 appears the main determinant of phenotype severity, with no evidence for a holoprosencephaly locus. J Med Genet. 2012;49(6):366–72.
- Devriendt K, Vanhole C, Matthijs G, de Zegher F. Deletion of thyroid transcription factor-1 gene in an infant with neonatal thyroid dysfunction and respiratory failure. N Engl J Med. 1998;338(18):1317–8.
- Stockton DW, Das P, Goldenberg M, D'Souza RN, Patel PI. Mutation of PAX9 is associated with oligodontia. Nat Genet. 2000;24(1):18–9.
- Villafuerte B, Natera-de-Benito D, Gonzalez A, Mori MA, Palomares M, Nevado J, et al. The brain-lung-thyroid syndrome (BLTS): a novel deletion in chromosome 14q13.2-q21.1 expands the phenotype to humoral immunodeficiency. Eur J Med Genet. 2018;61(7):393–8.
- Richards S, Aziz N, Bale S, Bick D, Das S, Gastier-Foster J, et al. Standards and guidelines for the interpretation of sequence variants: a joint consensus recommendation of the American College of Medical Genetics and Genomics and the Association for Molecular Pathology. Genet Med. 2015;17(5):405–24.
- Xie C, Tammi MT. CNV-seq, a new method to detect copy number variation using high-throughput sequencing. BMC Bioinformatics. 2009;10:80.
- Kearney HM, Thorland EC, Brown KK, Quintero-Rivera F, South ST. Working Group of the American College of medical genetics laboratory quality assurance C. American College of Medical Genetics standards and guidelines for interpretation and reporting of postnatal constitutional copy number variants. Genet Med. 2011;13(7):680–5.
- Kortum F, Das S, Flindt M, Morris-Rosendahl DJ, Stefanova I, Goldstein A, et al. The core FOXG1 syndrome phenotype consists of postnatal microcephaly, severe mental retardation, absent language, dyskinesia, and corpus callosum hypogenesis. J Med Genet. 2011;48(6):396–406.
- Ibn-Salem J, Kohler S, Love MI, Chung HR, Huang N, Hurles ME, et al. Deletions of chromosomal regulatory boundaries are associated with congenital disease. Genome Biol. 2014;15(9):423.
- Ellaway CJ, Ho G, Bettella E, Knapman A, Collins F, Hackett A, et al. 14q12 microdeletions excluding FOXG1 give rise to a congenital variant Rett syndrome-like phenotype. Eur J Hum Genet. 2013;21(5):522–7.
- Cellini E, Vignoli A, Pisano T, Falchi M, Molinaro A, Accorsi P, et al. The hyperkinetic movement disorder of FOXG1-related epileptic-dyskinetic encephalopathy. Dev Med Child Neurol. 2016;58(1):93–7.
- Allou L, Lambert L, Amsallem D, Bieth E, Edery P, Destree A, et al. 14q12 and severe Rett-like phenotypes: new clinical insights and physical mapping of FOXG1-regulatory elements. Eur J Hum Genet. 2012;20(12):1216–23.
- Dale RC, Grattan-Smith P, Nicholson M, Peters GB. Microdeletions detected using chromosome microarray in children with suspected genetic movement disorders: a single-Centre study. Dev Med Child Neurol. 2012;54(7):618–23.
- Gras D, Jonard L, Roze E, Chantot-Bastaraud S, Koht J, Motte J, et al. Benign hereditary chorea: phenotype, prognosis, therapeutic outcome and long term follow-up in a large series with new mutations in the TITF1/NKX2-1 gene. J Neurol Neurosurg Psychiatry. 2012;83(10):956–62.
- Peall KJ, Lumsden D, Kneen R, Madhu R, Peake D, Gibbon F, et al. Benign hereditary chorea related to NKX2.1: expansion of the genotypic and phenotypic spectrum. Dev Med Child Neurol. 2014;56(7):642–8.

- Hayashi S, Yagi M, Morisaki I, Inazawa J. Identical deletion at 14q13.3 including PAX9 and NKX2-1 in siblings from mosaicism of unaffected parent. J Hum Genet. 2015;60(4):203–6.
- Das P, Stockton DW, Bauer C, Shaffer LG, D'Souza RN, Wright T, et al. Haploinsufficiency of PAX9 is associated with autosomal dominant hypodontia. Hum Genet. 2002;110(4):371–6.
- Brunskill EW, Ehrman LA, Williams MT, Klanke J, Hammer D, Schaefer TL, et al. Abnormal neurodevelopment, neurosignaling and behaviour in Npas3-deficient mice. Eur J Neurosci. 2005;22(6):1265–76.
- Piccione M, Serra G, Consiglio V, Di Fiore A, Cavani S, Grasso M, et al. 14q13.
 1-21.1 deletion encompassing the HPE8 locus in an adolescent with intellectual disability and bilateral microphthalmia, but without holoprosencephaly. Am J Med Genet A. 2012;158A(6):1427–33.
- 24. Shimojima K, Komoike Y, Tohyama J, Takahashi S, Paez MT, Nakagawa E, et al. TULIP1 (RALGAPA1) haploinsufficiency with brain development delay. Genomics. 2009;94(6):414–22.
- Zhu M, Tao J, Vasievich MP, Wei W, Zhu G, Khoriaty RN, et al. Neural tube opening and abnormal extraembryonic membrane development in SEC23A deficient mice. Sci Rep. 2015;5:15471.
- Boyadjiev SA, Kim SD, Hata A, Haldeman-Englert C, Zackai EH, Naydenov C, et al. Cranio-lenticulo-sutural dysplasia associated with defects in collagen secretion. Clin Genet. 2011;80(2):169–76.
- Caliebe A, Martin Subero JI, Muhle H, Gesk S, Janig U, Krause M, et al. A 2 Mb deletion in 14q13 associated with severe developmental delay and hemophagocytic lymphohistiocytosis. Eur J Med Genet. 2011;54(5):e505–9.
- Kharbanda M, Hermanns P, Jones J, Pohlenz J, Horrocks I, Donaldson M. A further case of brain-lung-thyroid syndrome with deletion proximal to NKX2-1. Fur J Med Genet. 2017;60(5):257–60.
- Thorwarth A, Schnittert-Hubener S, Schrumpf P, Muller I, Jyrch S, Dame C, et al. Comprehensive genotyping and clinical characterisation reveal 27 novel NKX2-1 mutations and expand the phenotypic spectrum. J Med Genet. 2014;51(6):375–87.
- Petersheim D, Massaad MJ, Lee S, Scarselli A, Cancrini C, Moriya K, et al. Mechanisms of genotype-phenotype correlation in autosomal dominant anhidrotic ectodermal dysplasia with immune deficiency. J Allergy Clin Immunol. 2018;141(3):1060–73 e3.
- Ozaki K, Sato H, Iida A, Mizuno H, Nakamura T, Miyamoto Y, et al. A functional SNP in PSMA6 confers risk of myocardial infarction in the Japanese population. Nat Genet. 2006;38(8):921–5.
- Sukhov A, Adamopoulos IE, Maverakis E. Interactions of the immune system with skin and bone tissue in psoriatic arthritis: a comprehensive review. Clin Rev Allergy Immunol. 2016;51(1):87–99.
- Sjakste T, Paramonova N, Rumba-Rozenfelde I, Trapina I, Sugoka O, Sjakste N. Juvenile idiopathic arthritis subtype- and sex-specific associations with genetic variants in the PSMA6/PSMC6/PSMA3 gene cluster. Pediatr Neonatol. 2014;55(5):393–403.
- Conrad DF, Pinto D, Redon R, Feuk L, Gokcumen O, Zhang Y, et al. Origins and functional impact of copy number variation in the human genome. Nature. 2010;464(7289):704–12.
- Magi A, Tattini L, Cifola I, D'Aurizio R, Benelli M, Mangano E, et al. EXCAVATOR: detecting copy number variants from whole-exome sequencing data. Genome Biol. 2013;14(10):R120.
- Marchuk DS, Crooks K, Strande N, Kaiser-Rogers K, Milko LV, Brandt A, et al. Increasing the diagnostic yield of exome sequencing by copy number variant analysis. PLoS One. 2018;13(12):e0209185.
- Yao R, Zhang C, Yu T, Li N, Hu X, Wang X, et al. Evaluation of three readdepth based CNV detection tools using whole-exome sequencing data. Mol Cytogenet. 2017;10:30.
- Gao C, Wang X, Mei S, Li D, Duan J, Zhang P, et al. Diagnostic yields of trio-WES accompanied by CNVseq for rare neurodevelopmental disorders. Front Genet. 2019;10:485.
- Jiao Q, Sun H, Zhang H, Wang R, Li S, Sun D, et al. The combination of whole-exome sequencing and copy number variation sequencing enables the diagnosis of rare neurological disorders. Clin Genet. 2019;96(2):140–50.

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